



(19) Europäisches Patentamt
 European Patent Office
 Office européen des brevets



(11) Publication number:

0 408 770 A1

(22)

EUROPEAN PATENT APPLICATION
 published in accordance with Art.
 158(3) EPC

(21) Application number: 90902694.0

(51) Int. Cl. 5: C08B 37/00, A61K 31/725

(22) Date of filing: 06.02.90

(86) International application number:
 PCT/JP90/00141(87) International publication number:
 WO 90/08784 (09.08.90 90/19)

(30) Priority: 06.02.89 JP 28299/89

(43) Date of publication of application:
 23.01.91 Bulletin 91/04(84) Designated Contracting States:
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(54) NEW SULFATED POLYSACCHARIDE, PHARMACEUTICALLY ACCEPTABLE SALTS THEREOF,
 PREPARATION THEREOF, AND DRUG CONTAINING THE SAME AS ACTIVE INGREDIENT.

(57)

(α) 20° - - 55 ~ - 73° (C = 1%) (α)

This invention relates to a sulfated polysaccharide D-HG, prepared by depolymerizing FGAG (an extract from the parietes of a sea cucumber comprising a sulfated polysaccharide having a heparinlike action) or its salt and having the following physicochemical properties, pharmaceutically acceptable salts thereof, a method for preparing the same, and drugs for treating diffuse intravascular coagulation (DIC) and thrombosis containing the same as the active ingredient: (1) molecular weight: 3000 to 42000 according to HP GPC; (2) form: white, amorphous and highly hygroscopic powder; (3) solubility: soluble in water and insoluble in organic solvents such as ethanol and acetone; (4) specific rotation: (a); (5) color reaction: Elson-Morgan reaction (+); carbozole-sulfuric acid reaction (+); cysteine-sulfuric acid reaction (+); orcinol-hydrochloric acid reaction (+); azure A metachromasia reaction (+); (6) compositional analysis: galactosamine : glucuronic acid : fucose : sulfate radical = 1 : 0.8 ± 0.2 : 0.85 ± 0.15 : 3.4 ± 0.9.

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Specification

NOVEL SULFATED POLYSACCHARIDE, PHARMACEUTICALLY
ACCEPTABLE SALT THEREOF, PROCESS FOR PREPARING SAME AND
MEDICAMENT CONTAINING SAME AS EFFECTIVE COMPONENT

5

Technical Field

The present invention relates to a novel sulfated polysaccharide, a pharmaceutically acceptable salt thereof, a process for preparing the same and a medicament containing the same as an effective component.

10

Background Art

The present inventors separated a sulfated polysaccharide from the body wall of a sea cucumber by extraction with alkali, the sulfated polysaccharide having an anti-coagulation activity and a lipid clearing activity which are typical of heparin. The inventors named the polysaccharide FGAG (Yao Hsueh 1980, 15(5), 263-270, Zhongyao Tongbao 1982, 7(4), 27-29, Hsueh Pao 1983, 18(3), 203-208 and Japanese Unexamined Patent Publication No.63-128001). Japanese Unexamined Patent Publication No.63-10601 discloses another example of the separation of sulfated polysaccharide by other researchers. Although differently named, the sulfated polysaccharides described in the above prior art publications are all identical and have the following physicochemical constants.

25

Characteristic:

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white, amorphous, highly hygroscopic powder

Molecular weight:

about 15,000 to about 80,000 (as
measured by gel filtration)

5 Analysis for composition: As shown below.

Galactosamine	13 to 17 wt.%
Glucuronic acid	16 to 19 wt.%
Fucose	13 to 27 wt.%
Sulfate	27 to 38.5 wt.%

10 Molar ratio: As shown below.

Galactosamine: glucuronic acid: fucose:
sulfate = 1 : 1±0.2 : 1.35±0.35 : 3.6±0.6

According to the above analytical values and the
like, FGAG is identified as a high-molecular weight
15 sulfated polysaccharide comprising galactosamine,
glucuronic acid, fucose, etc. and is characterized by a
larger content of sulfate than known natural sulfated
polysaccharides.

With a high anti-coagulation activity, said FGAG
20 was once a candidate for a medicament for curing dis-
seminated intravascular coagulation (DIC). However, FGAG
was later found to have a high activity to cause platelet
aggregation when used for human beings and to be useless
in treatment of humans' DIC if used as it is because of
25 such side effect.

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Disclosure of the Invention

In view of the above situation, the present inventors conducted extensive research on compounds which can be used as an excellent medicament for treatment of DIC and which have activities like heparin's. Our finding was that the sulfated polysaccharide prepared by depolymerization of FGAG or a salt thereof exhibits substantially no activity to cause platelet aggregation while sustaining the anti-coagulation activity and other activities like heparin's. We further discovered that unlike heparin, the sulfated polysaccharide show an activity to inhibit the production of thrombin without displaying an anti-Xa or anti-thrombin activity, and thus may be potentially effective in treatment of thrombosis. The present inventors named the novel sulfated polysaccharide D-HG.

The present invention has been accomplished based on these novel findings.

According to the invention, there are provided a novel sulfated polysaccharide (D-HG), and a pharmaceutically acceptable salt thereof, a process for preparing the same and a medicament for treatment of DIC and thrombosis containing the above as the effective component.

D-HG of the invention has the physicochemical

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properties shown below.

[1] Molecular weight:

3,000 to 42,000 (as measured by high performance
GPC)

5 [2] Characteristic:

white, amorphous, highly hygroscopic powder

[3] Solubility:

soluble in water but insoluble in ethanol,
acetone and like organic solvents.

10 [4] Specific rotation:

$[\alpha]_D^{20} = -55 \text{ to } -73^\circ$ (C = 1%)

[5] Color reaction: As shown below

Elson-Morgan reaction +

Carbazole-sulfuric acid reaction +

15 Cysteine-sulfuric acid reaction +

Orcinol-hydrochloric acid reaction +

Azure A metachromasia reaction +

[6] Analysis for composition:

20 D-HG comprises constituent saccharides including
galactosamine (abbreviated to GalN), glucuronic acid
(abbreviated to GA) and fucose (abbreviated to Fuc)
and sulfate in a molar ratio of GalN : GA : Fuc :
sulfate = 1 : 0.8 ± 0.2 : 0.85 ± 0.15 : 3.4 ± 0.9.

Analyses were conducted by the following methods to
25 check galactosamine, glucuronic acid, fucose and sulfate.

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GalN:

White method (Carbohydrate Research, 114: 586, 201)

GA:

Bitter-Muir method (Anal. Biochem., 4: 330, 1962)

5

Fuc:

Dische method (J. Biol. Chem., 175: 595, 1948)

Sulfate:

Dodgson & Price method (Biochem. J., 84: 106, 1962)

The above analytic results show that D-HG has in the
10 molecule sulfate and carboxyl group which react with bases to
form a salt. D-HG is stable in the form of a salt and
isolated and purified usually in the form of a salt. Usable
as salts are pharmaceutically acceptable salts including salts of
15 potassium, sodium or like alkali metals, and salts of
calcium, magnesium, barium or like alkaline earth metals, or
pyridinium salt or like organic bases. Shown below is the
composition of constituent saccharides in a form in which a
salt is not formed, i.e., in free form.

	GalN	18 to 24 wt.%
20	GA	14 to 21 wt.%
	Fuc	13 to 20 wt.%
	Sulfate	31 to 44 wt.%

A preferred molecular weight of D-HG and a salt
thereof is about 4,000 to about 15,000 (as determined by
25 high performance GPC).

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D-HG of the invention is prepared from FGAG as a starting material. For preparation of D-HG, FGAG or a salt thereof is depolymerized, followed by isolation and purification. FGAG is obtained by decomposing the body wall of a sea cucumber, an oceanic life, with alkali, and further decomposing the resulting product with pancreatin or like proteolytic enzymes for extraction, followed by isolation and purification.

FGAG or a salt thereof for use in the preparation of D-HG of the invention can be easily produced by the methods disclosed in the known publications cited above in reference to the prior art, more specifically for example by the method to be described later in Reference Example. Examples of sea cucumbers useful in the preparation of FGAG or a salt thereof are:

Stichopus japonicus Selenka,
Stichopus chloronoyus Brandt,
Stichopus variegatus Semper,
Holothuria pervicax Selenka,
Holothuria atra,
Holothuria argus,
Holothuria edulis,
Holothuria scabra,
Parastichopus nigripunctatus,

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Thelenota ananas,
Holothuria monacaria Lesson,
Holothuria leucospilota Brandt,
Cucumaria chronhjelmi,
5 Cucumaria echinata,
Cucumaria frondosa Japonica,
Pentacta australis,
Paracaudina chilensis ransonneti,
Molpadia musculus,
10 Leptosynapta inhaerens,
Polycheira rufescens,
Synapta maculata,
Halodeima cinerascens (Brandt),
Actinopyga lacanora (Jaeger),
15 Actinopyga echinates (Jaeger),
Microthelie nobilis (Selenka), etc.
The sea cucumber to be used as the starting material may
be a raw or dried one. Of the sea cucumbers exemplified
above, Stichopus japonicus Selenka is most preferred as
20 the starting material.

D-HG is prepared by dissolving the above-obtained FGAG or a salt thereof in water and depolymerizing the solution. In the depolymerization reaction, a high-molecular weight sulfated polysaccharide such as heparin or the like is converted into a low-molecular
25

weight sulfated polysaccharide. A depolymerizing agent is usually used in the reaction. Examples of useful depolymerizing agents are hydrogen peroxide, hypochlorous acid, hypobromous acid, sodium hypochlorate and like 5 hypohalogenous acids and salts thereof; periodic acid, sodium periodate and like periodic acids and salts thereof, etc. Ascorbic acid, ferrous ion or the like is usable as a reaction accelerator. Optionally, the depolymerization reaction may be effected by application 10 of radiations such as ultrasonic waves, ultraviolet rays, gamma rays or the like alone in lieu of a depolymerizing agent or in combination with the above depolymerizing agent. The most preferred depolymerization method in the invention is one using hydrogen peroxide as a 15 depolymerizing agent. Hydrogen peroxide is reacted in an amount of 1 to 31 wt.%, preferably 1 to 16 wt.% in terms of a hydrogen peroxide concentration. The reaction time is usually 1 to 60 hours, preferably 3 to 40 hours, and the reaction temperature ranges from room temperature to 20 about 80°C, preferably about 40 to about 60°C. The pH range in the reaction of hydrogen peroxide is acidic or neutral in the range of from 1 to 8, preferably 3 to 7. To maintain a constant pH value, hydrogen peroxide may be reacted in a buffer such as acetate buffer, phosphate 25 buffer, Tris buffer or the like, or a pH controller using

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diluted sodium hydroxide or the like may be used in the reaction. On completion of the reaction, the pH is returned to neutral range, and isolation and purification are conducted. The isolation and purification can be
5 done, for example, by fractional precipitation using an organic solvent such as ethanol, acetone or the like; acetate such as potassium acetate, barium acetate, calcium acetate, ammonium acetate or the like; or quaternary ammonium salt such as cetyltrimethyl ammonium salt or the like. The isolation and purification is also feasible by
10 ion exchange chromatography using resins such as DEAE-Cellulose (product of Sigma Chemical Co.), DEAE-Toyopearl (product of Tosoh Corporation), DEAE-Cellulofine (product of Chisso Corporation), Dowex-1 (product of Dow Chemical Co.) or the like, or gel filtration chromatography using resins such as Sephadex G-50, Sephadex G-200 (both products of Pharmacia-LKB Biotechnology), by dialysis using Spectra/Por (product of Spectrum Medical Industries, Inc.) or by ultrafiltration. These means are employed
15 alone or in a suitable combination thereof. Ion exchange chromatography, gel filtration chromatography and ultrafiltration are preferable to produce easily D-HG having no activity to cause platelet aggregation.
20

The thus obtained D-HG is usually isolated in
25 the form of a salt of sodium and/or potassium or the

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like. D-HG in a salt form can be transformed into free D-HG by treatment with a cation-exchange resin such as Dowex 50W or the like. D-HG in a salt form, if necessary, can be converted into a desired pharmaceutically acceptable salt by salt exchange commonly employed. Usable as salts of sulfated polysaccharides are pharmaceutically acceptable salts including salts of potassium, sodium or like alkali metals, and salts of calcium, magnesium, barium or like alkaline earth metals, or pyridinium salt or like organic bases.

The treatment of DIC and thrombosis by D-HG of the invention is conducted utilizing its anti-coagulation activity against the acceleration of coagulation in blood vessels which causes DIC and thrombosis. The range of anti-coagulation activity of D-HG includes an activity to inhibit the platelet aggregation by thrombin as well as an anti-coagulation enzyme activity, typically an activity to prolong the activated partial thromboplastin time. The anti-coagulation activity of D-HG is entirely different from heparin's in that D-HG does not require any plasma factor such as anti-thrombin III in exhibiting the activity nor is influenced by the anti-heparin factor such as platelet factor 4. A further difference from heparin is that D-HG has an activity to inhibit the production of thrombin without displaying an anti-Xa activity or an

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anti-thrombin activity, hence apparently effective against thrombosis. The feature of D-HG is that unlike heparin and FGAG, D-HG has substantially no activity to cause platelet coagulation, which is the fatal activity of medicaments for treatment of DIC and thrombosis. The expression "substantially no activity to cause platelet coagulation" means that when administered to organisms, especially human beings, D-HG does not exhibit platelet coagulation which poisons organisms or aggravates thrombosis.

D-HG is made into various pharmaceutical compositions useful for DIC and thrombosis treatment. Stated more specifically, the composition comprising an effective amount of D-HG and/or a pharmaceutically acceptable salt and a pharmaceutically acceptable carrier can be prepared in various administration forms. The administration form can be any of tablets, capsules, powders, granules, grains, solutions, emulsions, suspensions and like oral forms, and injections, suppositories, ointment, plaster and like parenteral forms. These preparations can be manufactured by conventional methods already known to those skilled in the art. A solid preparation for oral administration can be prepared by mixing the effective component of the invention with an excipient with or without addition of

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binders, disintegrators, lubricants, coloring agents, flavorings, perfumes, etc. and making the mixture into tablets, capsules, powders, granules, grains or the like in a conventional manner. Injection preparations can be
5 produced by adding a pH-adjusting agent, buffer, stabilizer, isotonizing agent, local anesthetic and the like to the effective component, and making the mixture into intravenous, intramuscular, subcutaneous, intracutaneous or intraperitoneal injections in a
10 conventional manner. Suppositories can be prepared by making a mixture of the effective component, base materials and optionally a surfactant and the like into a suppository in a conventional manner.

Examples of excipients useful for oral solid preparations are lactose, sucrose, starch, talc, magnesium stearate, crystalline cellulose, methyl cellulose, carboxymethyl cellulose, glycerin, sodium alginate, gum arabic, etc. Examples of binders useful for oral preparations include polyvinyl alcohol, polyvinyl ether,
15 ethyl cellulose, gum arabic, shellac, sucrose, etc. Examples of useful lubricants are magnesium stearate, talc and the like. The coloring agents, disintegrators and other auxiliaries to be added include those commonly used in the art. Tablets may be coated by well-known methods.
20

25 Examples of base materials useful for

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suppositories include oily base materials such as macrogol, lanolin, cacao oil, fatty acid triglyceride, Witepsol (registered trademark for the product of Dynamite Nobel) and so on.

5 The amount of the effective component per each unit dosage varies with the symptoms of the patient to be given the preparation, the form of the preparation, etc. Usually a preferred amount is 10 to 200 mg in an oral preparation, 1 to 100 mg in an injection, or 10 to 100 mg. 10 in a suppository, per unit dosage. The daily clinical dosage of the composition of the invention also varies with the patient's age, sex, conditions and other factors but usually may be in the range of about 10 to about 1,000 mg, preferably about 50 to about 200 mg in terms of the 15 effective component and can be given at 1 to 4 divided doses.

According to the present invention, there is provided a novel sulfated polysaccharide, D-HG, having substantially no activity to cause platelet coagulation and having an excellent anti-coagulation activity and remarkable characteristics as a medicament for treatment 20 of DIC and thrombosis.

Best Mode for Carrying Out the Invention

25 The present invention will be described in greater detail with reference to Reference Example,

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Examples and Pharmacological Tests. The percentages in Reference Example and Examples are all by weight.

Reference Example 1

Preparation of FGAG

5 One kilogram of dried *Stichopus japonicus* was immersed in 10 l of warm water and left to swell overnight. The flesh was removed and homogenized. Potassium hydroxide was added in an amount to give a 1N mixture. The mixture was treated at 60°C for 100 minutes
10 and adjusted to a pH of 8.5. After addition of 50 g of pancreatin, the mixture was stirred at 50°C for 3 hours.

After removal of impurities by centrifugation, 4.3 l of ethanol was added to the residue. The mixture was allowed to stand at 4°C and the resulting precipitate
15 was collected. The precipitate was washed with 80% ethanol, anhydrous ethanol and acetone in this sequence, and dried under reduced pressure, giving 50 g of a crude product. Fifty grams of the crude product was dissolved in 3.5 l of water and the solution was centrifuged to remove the insolubles. To the supernatant were added 5% sodium chloride and 40% ethanol to give a precipitate.
20 The precipitate was collected by centrifugation. After the precipitate was dissolved in 2.5 l of water, the solution was adjusted to a pH of 10.5. To the solution
25 was added dropwise a 30% aqueous solution of hydrogen

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peroxide. The mixture was decolorized in a water bath at 50°C with heating for about 3 hours. After cooling, the insolubles were removed by centrifugation. To the supernatant was added about 490 g of potassium acetate and 5 the mixture was kept at 4°C overnight. The following day, the resulting precipitate was dissolved in 2 l of water, the solution was cooled to 0°C, and the pH was adjusted to 2.8. The insolubles were removed from the solution by centrifugation. After neutralizing the supernatant, 196 g 10 of potassium acetate was added. The mixture was allowed to stand at 4°C to give a precipitate, which was then collected by centrifugation. The precipitate was dissolved in water to give a solution having a potassium acetate concentration of 0.5M and the solution was left 15 overnight at 4°C. The precipitate was collected by centrifugation, washed with 40% methanol and dissolved in 1 l of water. To the solution were added 5% sodium chloride and 40% ethanol to give a precipitate. The precipitate was collected by centrifugation, washed with 20 80% methanol, anhydrous ethanol and acetone in this sequence, and dried under reduced pressure, giving 17 g of a FGAG sodium/potassium salt. The physicochemical constants of the salt are as follows.

Molecular weight:

25 55,000 (as determined by high performance GPC)

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Analysis for composition: As shown below

	GalN:	20.0 %
	GA:	18.6 %
	Fuc:	17.2 %
5	Sulfate:	36.6 %
	Na:	6.2 %
	K:	7.4 %

Example 1

Ten grams of the FGAG sodium/potassium salt
10 obtained in Reference Example 1 was dissolved in 75 ml of water and 25 ml of a 30% aqueous solution of hydrogen peroxide was added. While maintaining the solution at a pH of about 7 with a diluted sodium hydroxide solution using a pH controller, the solution was heated at 60°C for
15 12 hours. After cooling, 2% sodium chloride and 40% ethanol were added to give a precipitate. The precipitate was collected by centrifugation, washed with 80% ethanol, anhydrous ethanol and acetone in this sequence, and dried under reduced pressure, giving 7.15 g of a D-HG
20 sodium/potassium salt.

Example 2

A 6.95 g quantity of a D-HG sodium/potassium salt was prepared by the same procedure as in Example 1 with the exception of treatment with hydrogen peroxide for
25 24 hours.

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Example 3

A D-HG sodium/potassium salt was prepared by the same procedure as in Example 1 with the exception of conducting the reaction while maintaining the pH of about 5 4 with a diluted alkali solution. Yield 6.4 g.

Example 4

Ten grams of the FGAG sodium/potassium salt obtained in Reference Example 1 was dissolved in 83.3 ml of a 0.2M phosphate buffer (pH 7.0). To the solution was 10 added 16.7 ml of a 30% aqueous solution of hydrogen peroxide. The mixture was treated at 60°C for 12 hours. After cooling, 2% sodium chloride and 40% ethanol were added to give a precipitate. The precipitate was collected by centrifugation, washed with 80% ethanol, 15 anhydrous ethanol and acetone in this sequence, and dried under reduced pressure, giving 7.18 g of a D-HG sodium/potassium salt.

Example 5

A D-HG sodium/potassium salt was prepared by the 20 same procedure as in Example 4 with the exception of conducting the reaction using a 0.2M acetate buffer (pH 3.5). Yield 7.05 g.

Examples 6 and 7

Two grams of the FGAG sodium/potassium salt 25 obtained in Reference Example 1 was dissolved in 15 ml of

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water. To the solution was added 5 ml of a 30% aqueous solution of hydrogen peroxide, and the mixture was treated at 60°C for 14 hours (Example 6) or 40 hours (Example 7). After cooling, the mixture was adjusted to a pH of 7 to 8 and thoroughly dialyzed against water using Spectra/por 3. The mixture was lyophilized and dried under reduced pressure. In this way, 1.62 g and 1.76 g of D-HG sodium/potassium salts were prepared.

Example 8

10 Two grams of the FGAG sodium/potassium salt obtained in Reference Example 1 was dissolved in 16.7 ml of water. To the solution was added 3.3 ml of a 30% aqueous solution of hydrogen peroxide, and the mixture was treated at 45°C for 24 hours. After cooling, the mixture 15 was returned to a pH of about 7 after which 2% sodium chloride and 40% ethanol were added to provide a precipitate. The precipitate was collected by centrifugation, washed with 80% ethanol, anhydrous ethanol and acetone in this sequence, and dried under reduced 20 pressure, giving 1.41 g of a D-HG sodium/potassium salt.

Examples 9 to 12

Two grams of the FGAG sodium/potassium salt obtained in Reference Example 1 was dissolved in 15 ml of water. To the solution was added 5 ml of a 30% aqueous 25 solution of hydrogen peroxide, and the mixture was treated

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at 60°C for 4, 8, 12 or 24 hours. After cooling, the mixture was adjusted to a pH of 7 to 8 and fully dialyzed against water using Spectra/por 3. The same treatment as in Example 8 followed. In this way, 1.42 g, 1.35 g, 1.35 g and 1.2 g of D-HG sodium/potassium salts were produced.

Examples 13 and 14

Two grams of the FGAG sodium/potassium salt obtained in Reference Example 1 was dissolved in 14.7 ml of water. To the solution was added 5.3 ml of a 30% aqueous solution of hydrogen peroxide. The mixture was treated at 45°C for 14 hours or 40 hours. After cooling, the mixture was returned to a pH of about 7 after which 2% sodium chloride and 40% ethanol were added to give a precipitate. The same treatment as in Examples 6 to 7 followed. In this way, 1.64 g and 1.62 g of D-HG sodium/potassium salts were prepared.

Example 15

Two grams of the FGAG sodium/potassium salt obtained in Reference Example 1 was dissolved in 30 ml of water and treated for 12 hours in the same manner as in Example 8. The solution was fractionated with a solution of sodium chloride on a Sephadex G-50T column (product of Pharmacia-LKB Biotechnology). While monitoring uronic acid, peaks were divided into three. The eluate obtained last was collected, fully dialyzed against water,

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lyophilized and dried under reduced pressure, giving 0.2 g of a D-HG sodium salt.

Example 16

5 A 0.5 g quantity of the D-HG sodium/potassium salt obtained in Example 8 was fractionated in the same manner as in Example 15, giving 0.18 g of a D-HG sodium salt.

10 Fig. 1 shows an infrared absorption spectrum of the D-HG sodium salt (as measured with KBr tablet) and Fig. 2 a proton nuclear magnetic resonance spectrum (in D₂O, 90MHz, 70°C) thereof.

Example 17

15 Two grams of the FGAG sodium/potassium salt obtained in Reference Example 1 was dissolved in 16.7 ml of water, and 3.3 ml of a 30% aqueous solution of hydrogen peroxide was added. The mixture was treated at 45°C for 5 hours. After cooling, the mixture was returned to a pH of about 7 after which 2% sodium chloride and 40% ethanol were added to give a precipitate. The precipitate was 20 collected by centrifugation, washed with 80% ethanol, anhydrous ethanol and acetone in this sequence, and dried under reduced pressure, giving a D-HG sodium/potassium salt. Yield 1.60 g.

Example 18

25 Two grams of the FGAG sodium/potassium salt

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obtained in Reference Example 1 was treated for 9 hours by the same method as in Example 17, giving 1.54g of a D-HG sodium/potassium salt.

Example 19

5 Two grams of the FGAG sodium/potassium salt obtained in Reference Example 1 was treated for 12 hours by the same method as in Example 17, giving 1.52 g of a D-HG sodium/potassium salt.

Example 20

10 One gram of the FGAG sodium/potassium salt obtained in Reference Example 1 was dissolved in 8.7 ml of 0.2M phosphate buffer (pH 7.0). To the solution was added 1.3 ml of a 30% aqueous solution of hydrogen peroxide and the mixture was treated at 60°C for 3 hours. After
15 cooling, 5% sodium chloride and 66% ethanol were added to the mixture to give a precipitate. The precipitate was collected by centrifugation, washed with 80% ethanol, anhydrous ethanol and diethyl ether in this sequence, and dried under reduced pressure, giving a D-HG sodium/potassium salt. The obtained salt was dissolved in
20 10 ml of 20mM Tris-HCl buffer (pH 7.0) and the solution was admixed with DEAE-Toyopearl (product of Tosoh Corporation) thoroughly equilibrated with the buffer. Elution was conducted in the buffer with a linear
25 concentration gradient of sodium chloride (0 to 1 M).

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While monitoring uronic acid, peak fractions were collected and precipitation occurred with addition of a two-fold amount of ethanol. The precipitate was collected by centrifugation, washed with 80% ethanol, anhydrous ethanol and diethyl ether in this order and dried under reduced pressure, giving a D-HG sodium salt. Yield 0.39 g.

Table 1 shows physicochemical properties of D-HG's obtained in the above Examples.

Table 1

Ex.	MW x 10 ³	[α] _D ²⁰	Composition				Molar ratio	
			GaIN	FUC	GA	SuL*	Na	K
1	10.2	-71.2	21.7	16.7	16.5	29.7	7.1	6.4 1 : 0.70 : 0.84 : 2.5
2	9.7	-67.1	19.0	15.0	15.1	30.2	7.2	7.9 1 : 0.70 : 0.87 : 2.8
3	14.1	-71.5	20.2	17.6	15.6	33.1	6.2	6.7 1 : 0.71 : 0.95 : 3.0
4	4.7	-55.4	19.8	15.1	13.3	31.4	8.2	4.8 1 : 0.62 : 0.83 : 2.9
5	6.5	-61.2	20.6	15.6	15.8	32.8	7.0	4.4 1 : 0.71 : 0.83 : 2.9
6	12.0	-70.8	18.6	16.5	17.1	38.6	4.7	6.1 1 : 0.85 : 0.97 : 3.8
7	7.8	-68.0	17.5	13.1	16.1	37.1	5.4	6.4 1 : 0.85 : 0.82 : 3.9
8	13.4	-70.1	17.1	14.6	14.6	32.7	8.1	5.5 1 : 0.79 : 0.93 : 3.5
9	14.1	-69.6	19.9	14.3	14.8	34.0	7.6	5.2 1 : 0.69 : 0.79 : 3.1
10	8.6	-66.3	20.8	15.1	14.3	34.8	8.5	5.5 1 : 0.64 : 0.79 : 3.1

Table 1 (Continued)

Ex.	MW x 10 ⁻³	$[\alpha]_D^{20}$		Composition				Molar ratio	
		GaIN	Fuc	GA	Sul*	Na	K	GaIN : GA : Fuc : Sul*	
11	7.5	-66.0	17.8	13.2	32.9	6.4	5.1	1 : 0.68 : 0.81 : 3.4	
12	5.6	-64.8	20.1	14.7	33.4	4.1	5.1	1 : 0.60 : 0.80 : 3.0	
13	10.8	-72.2	18.1	14.8	16.7	37.1	5.0	6.5	1 : 0.85 : 0.89 : 3.8
14	6.6	-68.4	17.3	12.8	15.9	35.3	5.6	6.7	1 : 0.85 : 0.81 : 3.7
15	10.2	-68.2	19.1	15.8	14.4	32.9	8.6	0	1 : 0.69 : 0.90 : 3.1
16	7.7	-67.5	17.6	13.9	14.1	32.1	4.4	0	1 : 0.74 : 0.86 : 3.3
17	41.4	-70.7	19.8	14.7	15.8	29.9	5.9	5.5	1 : 0.73 : 0.81 : 2.7
18	24.3	-72.6	18.0	15.9	16.6	33.4	6.2	5.2	1 : 0.85 : 0.96 : 3.4
19	20.1	-71.1	18.3	14.4	14.8	33.0	7.5	5.0	1 : 0.75 : 0.86 : 3.3
20	12.8	-62.4	19.3	14.5	16.2	34.3	10.7	0	1 : 0.78 : 0.81 : 3.3

Note: In Table 1, the molecular weight (MW) was determined by high performance

GPC. Sul* stands for sulfate.

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D-HG's obtained in Examples 1 to 19 showed single spots in electrophoresis (Dietrich. C. P., J. Chromatogr., 130, 299 (1977)).

Preparation Example 1

5 Injection preparation

D-HG sodium salt prepared in Example 16 was dissolved in distilled water for injection to give a 5% aqueous solution. A 50 mg quantity (in terms of D-HG) of the solution was filled into a vial to perform lyophilization. A 2 ml quantity of physiological saline was added 10 as a solvent.

Preparation Example 2

Injection preparation

An injection preparation was prepared according 15 to the formulation as shown below.

D-HG sodium/potassium salt 40 mg
(Example 12)

Physiological saline q.s.

Per ampule 2 ml

Preparation Example 3

20 Tablet

Tablets were prepared according to the formulation as shown below.

D-HG sodium/potassium salt 10 mg
(Example 14)

25 Corn starch 65 mg

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Carboxymethyl cellulose	20 mg
Polyvinyl pyrrolidone	3 mg
<u>Magnesium stearate</u>	<u>2 mg</u>
Per tablet	100 mg

5

Preparation Example 4

Suppository

A suppository was prepared according to the formulation as shown below.

D-HG sodium/potassium salt (Example 4)	50 mg
Witepsol W-35 <u>(Product of Dynamite-Nobel AG)</u>	950 mg
Per suppository	1000 mg

Pharmacological Test

<Effect on DIC model>

15 D-HG, FGAG and heparin were tested for an effect on DIC model in accordance with the method described in Japan J. Pharmacol., 35, 203-227 (1984).

20 Used as D-HG was the sodium salt obtained in Example 16, as FGAG the sodium/potassium salt obtained in Reference Example 1 and as a heparin a sodium salt having a potency of 185.6 U/mg.

25 A 800 U/kg of thrombin was intravenously injected into ICR mice (10 to 15 mice a group). After 24 hours, the fatality of mice caused by DIC was observed to calculate the survival rate. The D-HG sodium salt, FGAG

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sodium/potassium salt or heparin sodium salt was intravenously injected one minute before the administration of thrombin. Table 2 shows the results.

Table 2

5

Medicament	Dose (mg/kg)	Survival rate (%)
Control	0	13
D-HG sodium salt	3	90
	1	60
	0.3	40
FGAG sodium/ potassium salt	1	60
<u>Heparin sodium salt</u>	<u>1</u>	<u>80</u>

10

15

20

D-HG produced the same anti-DIC effect as heparin and FGAG when used in an amount of 1 mg/kg. This model also serves as a thrombosis model, hence effective against thrombosis.

<Anti-coagulation Activity>

The D-HG sodium salt (Example 16) or D-HG sodium/potassium salt (Example 11) was added to citric acid-containing plasma obtained from a rabbit to a concentration of 10 µg/ml. The activity to prolong the activated partial thromboplastin time (APTT) against the control (physiological saline) was observed. Table 3 shows the results.

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Table 3

<u>Medicament</u>	<u>APTT (Δ sec)</u>
Ex. 16	24.7
Ex. 11	16.2
<u>Control</u>	<u>0.0</u>

5

D-HG exhibited a remarkable anticoagulation activity.

10

<Anti-coagulation Activity in Human>

Using the citric acid-containing plasma obtained from more than 6 normal persons, the D-HG sodium salt (Example 16), a FGAG sodium/potassium salt and a heparin sodium salt were each observed for the activity in respect of anticoagulation parameters ($\mu\text{g}/\text{ml}$). Table 4 shows the results.

15

$x2\text{APTT}$ shows the concentration ($\mu\text{g}/\text{ml}$) required to double the activated partial thromboplastin time of the control (without addition of a medicament).

20

IIaIC_{90} is a concentration ($\mu\text{g}/\text{ml}$) for 90% inhibition of thrombin activity which was calculated by measuring the activity to prolong the thrombin time.

25

XaIC_{50} is a medicament concentration ($\mu\text{g}/\text{ml}$) for 50% inhibition of decomposition of synthetic substrate S2222 with a factor X.

VIII-IC_{80} is a medicament concentration ($\mu\text{g}/\text{ml}$) for 80% inhibition of factor VIII which was calculated by

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measuring the activity to prolong the contact-activated coagulation time in the presence of a small amount of factor VIII using a factor VIII deficient plasma.

5 IIaGI is the concentration ($\mu\text{g}/\text{ml}$) required to double the control's time for complete inactivation of prothrombin in the contact-activated plasma. This represents an activity to inhibit the thrombin generation.

Table 4

	Medicament	$\times 2\text{APTT}$	IIaIC_{90}	XaIC_{50}	VIII IC_{80}	IIaGI
10	Heparin sodium salt	1.2	0.3	3.4	0.79	1.2
	FGAG sodium/potassium salt	2.4	2	5200	1.68	2.4
	Ex. 16	12.0	30	5100	4.77	12.0

15 As seen from the APTT prolonging activity, D-HG sodium salts have an anti-coagulation activity, but unlike heparin sodium salts, have substantially no anti-thrombin activity or anti-factor Xa activity. On the other hand, 20 D-HG sodium salts show an activity to inhibit the thrombin generation, which confirms that the salts have an anti-thrombosis activity. This activity is presumably due to the inhibition of factor VIII activity and the inhibition of positive feedback mechanism of coagulation cascade. The above indicate that D-HG is a remarkably 25 unique agent for treatment of DIC or thrombosis.

- 30 -

<Inhibitory Activity against Thrombin-induced Platelets Aggregation>

The result of addition of the D-HG sodium salt obtained in Example 16 or a heparin sodium salt having a potency of 185.6 U/mg was evaluated by observing the aggregation of platelets (expressed in an increase of light transmittance) caused by the addition of 0.1 U/ml of thrombin to a suspension of plasma-free washed platelet obtained from a rabbit. Table 5 shows the results.

10

Table 5

Medicament	Concentration (μ g/ml)	Light transmittance (%)
Control	0	86.6
D-HG sodium salt	3	46.4
	10	5.2
Heparin sodium salt	3	80.2

15

20

D-HG, unlike heparin, showed a remarkable inhibitory activity against thrombin aggregation in a plasma-free system. Thus, it was confirmed that the activity of D-HG is independent of the plasma factors such as ATIII, etc.

25

<Activity to Cause Platelets Aggregation in Human>

A citrated platelet-rich plasma was obtained from five normal persons (B, E, G, H, J). The D-HG sodium salt obtained in Example 16 or FGAG sodium/potassium salt

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obtained in Reference Example 1 was added to the plasma, and the resulting activity to cause platelet aggregation (expressed in an increase of light transmittance) was evaluated by observation. Table 6 shows the results.

5

Table 6

Medicament	Concentration (mg/ml)	Light transmittance (%)					J
		B	E	G	H		
D-HG sodium salt	1	1.2	2.5	3.8	3.0	1.3	
FGAG sodium/ potassium salt	0.3	20.9	83.8	48.2	65.4	77.4	
Control	-	2.4	1.3	2.3	1.8	2.4	

10

15

D-HG did not have an activity to cause platelet aggregation in a concentration of 1 mg/ml, but FGAG exhibited an activity to cause platelet aggregation in a lower concentration of 300 µg/ml.

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CLAIMS

1. A sulfated polysaccharide (D-HG) prepared by depolymerization of FGAG or a salt thereof and having the following physicochemical properties, and a pharmaceutically acceptable salt thereof:

5 [1] Molecular weight:

3,000 to 42,000 (as measured by high performance GPC)

[2] Characteristic:

10 white, amorphous, highly hygroscopic powder

[3] Solubility:

soluble in water but insoluble in ethanol, acetone and like organic solvents

[4] Specific rotation:

15 $[\alpha]_D^{20} = -55 \text{ to } -73^\circ (\text{C} = 1\%)$

[5] Color reaction: as shown below

	Elson-Morgan reaction	+
	Carbazole-sulfuric acid reaction	+
	Cysteine-sulfuric acid reaction	+
20	Orcinol-hydrochloric acid reaction	+
	Azure A metachromasia reaction	+

[6] Analysis for composition: as shown below

Galactosamine : Glucuronic acid : Fucose : sulfate =
1 : 0.8 ± 0.2 : 0.85 ± 0.15 : 3.4 ± 0.9.

25 2. A sulfated polysaccharide (D-HG) and a salt

thereof according to claim 1 which has a molecular weight of 4,000 to 15,000 (as measured by high performance GPC).

3. A process for preparing the sulfated polysaccharide (D-HG) and the salt thereof as defined in 5 claim 1, the process comprising the steps of depolymerizing FGAG or a salt thereof, and subjecting the resulting product to separation and purification.

4. A process according to claim 3 wherein the separation and purification are carried out by fractional 10 precipitation using potassium acetate and/or precipitation using ethanol.

5. A process according to claim 3 wherein the separation and purification are carried out by a gel filtration.

15 6. A process according to claim 3 wherein the separation and purification are carried out by an ion exchange.

7. A medicament for treatment of disseminated intravascular coagulation comprising an effective amount 20 of the sulfated polysaccharide (D-HG) and/or the salt thereof as defined in claim 1 and a pharmaceutically acceptable carrier.

8. A medicament for treatment of thrombosis comprising an effective amount of the sulfated 25 polysaccharide (D-HG) and/or the salt thereof as defined

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in claim 1 and a pharmaceutically acceptable carrier.

9. Use of the sulfated polysaccharide (D-HG) and/or the salt thereof as defined in claim 1 for preparation of a pharmacological composition for treatment of
5 disseminated intravascular coagulation syndrome.

10. Use of the sulfated polysaccharide (D-HG) and/or the salt thereof as defined in claim 1 for preparation of a pharmacological composition for treatment of thrombosis.

Regional phase of
TAIHO PHARMACEUTIC
KOTAI KASEI CO., I
Our ref.: A 434 EI

EP 0 408 770 A1

MOGENSEN & PARTNER
COPENHAGEN, DENMARK

CLAIMS SET FOR ES

5 1. A process for preparing a sulfated polysaccharide
(D-HG) having the following physicochemical properties:

[1] Molecular weight:

10 3,000 to 42,000 (as measured by high performance
GPC)

[2] Characteristic:

white, amorphous, highly hygroscopic powder

[3] Solubility:

15 soluble in water but insoluble in ethanol,
acetone and like organic solvents

[4] Specific rotation:

20 $[\alpha]_D^{20} = -55 \text{ to } -73^\circ$ (C = 1%)

[5] Color reaction: as shown below

Elson-Morgan reaction	+
Carbazole-sulfuric acid reaction	+
Cysteine-sulfuric acid reaction	+
Orcinol-hydrochloric acid reaction	+
Azure A metachromasia reaction	+

[6] Analysis for composition: as shown below

30 Galactosamine : Glucuronic acid : Fucose : sulfate =
1 : 0.8 ± 0.2 : 0.85 ± 0.15 : 3.4 ± 0.9

35 and a pharmaceutically acceptable salt thereof, comprising
the steps of depolymerizing FGAG or a salt thereof, and
subjecting the resulting product to separation and purifica-
tion.

2. A process according to claim 1 wherein the sulfated polysaccharide (D-HG) has the molecular weight of 4,000 to 15,000 (as measured by high performance GPC).

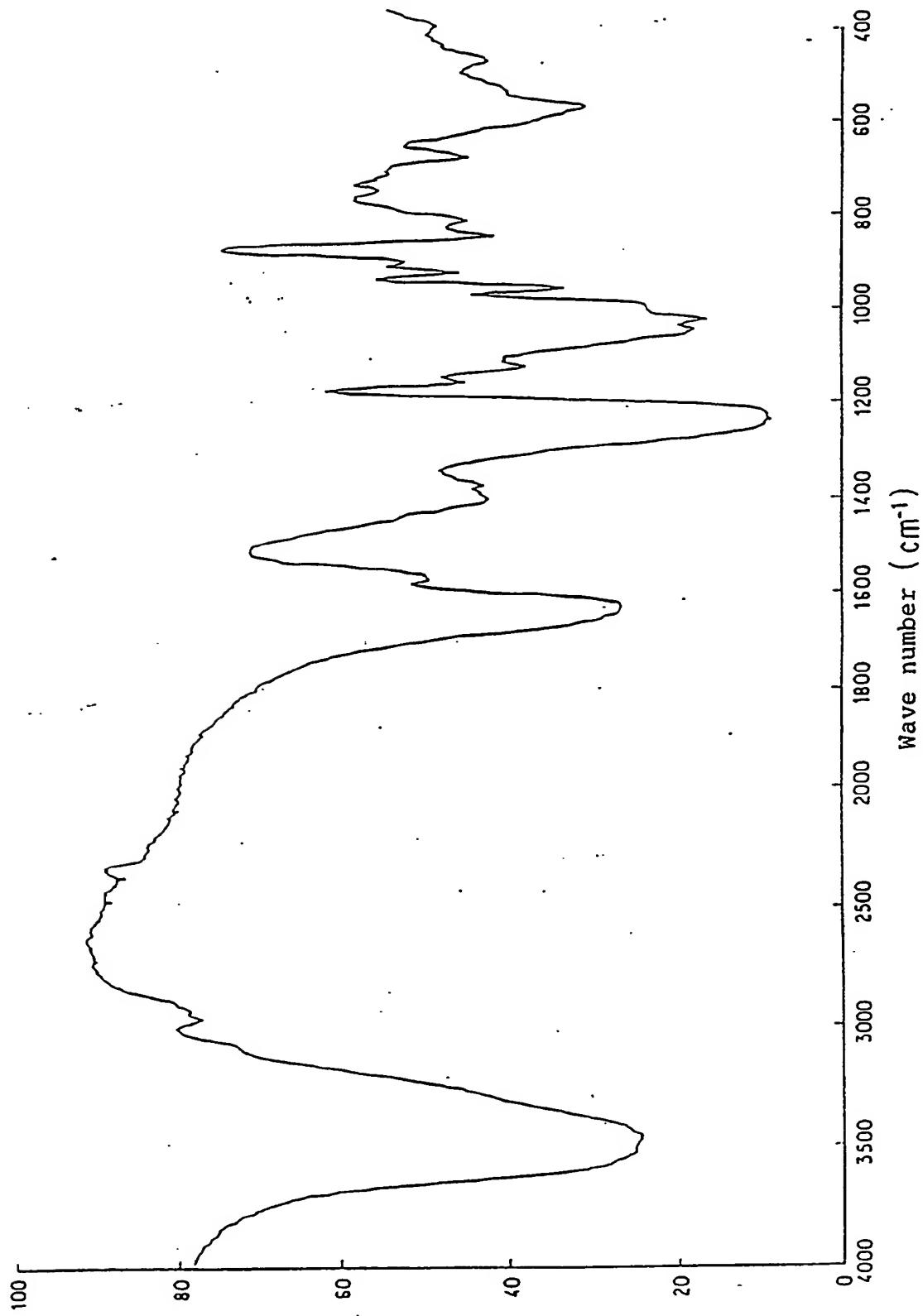
5 3. A process according to claim 1 or 2 wherein the separation and purification are carried out by fractional precipitation using potassium acetate and/or precipitation using ethanol.

10 4. A process according to claim 1 or 2 wherein the separation and purification are carried out by a gel filtration.

15 5. A process according to claim 1 or 2 wherein the separation and purification are carried out by an ion exchange.

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Fig. 1



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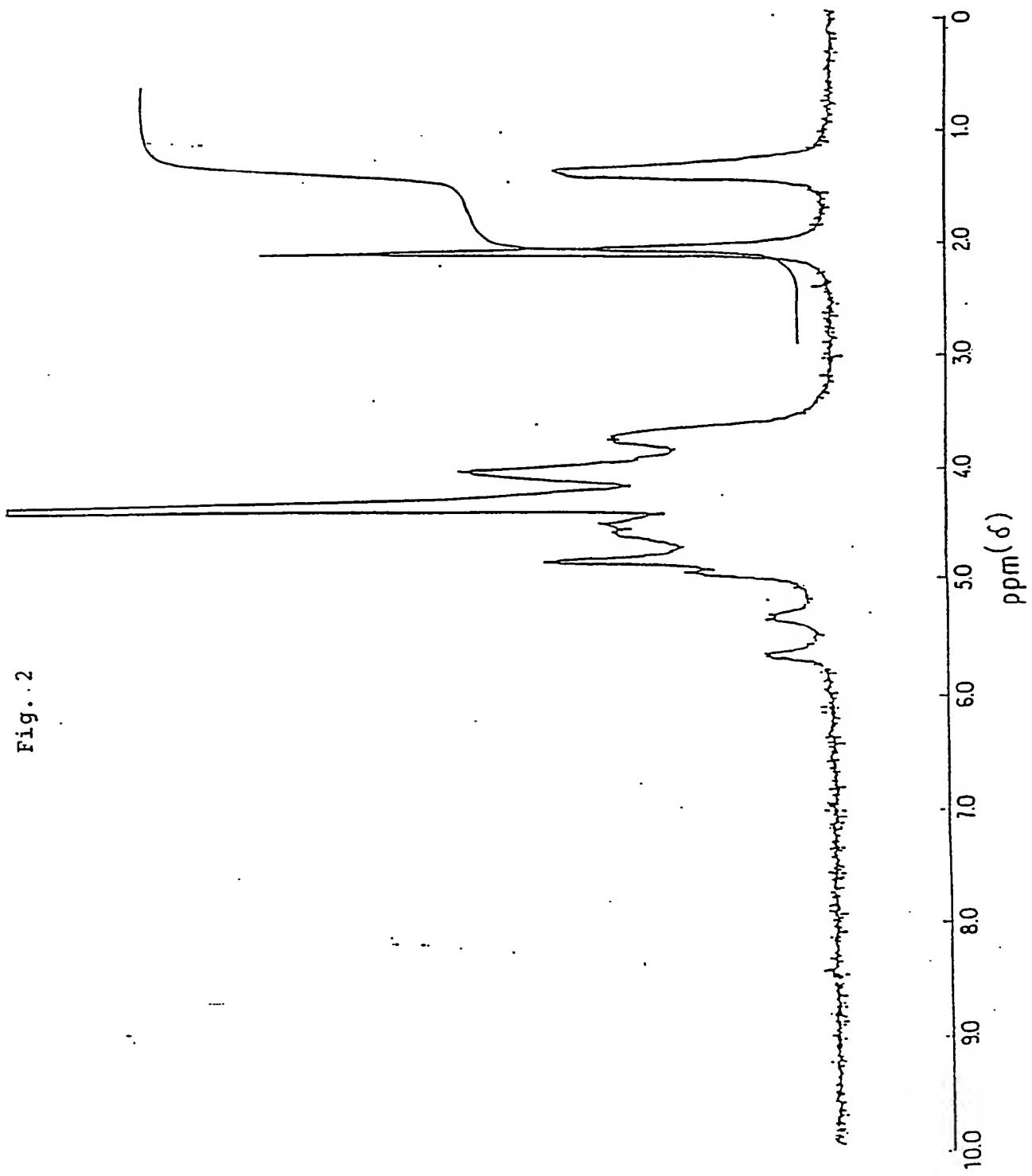


Fig. 2

INTERNATIONAL SEARCH REPORT

International Application No. PCT/JP90/00141

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all)¹

According to International Patent Classification (IPC) or to both National Classification and IPC

Int. Cl⁵ C08B37/00, A61K31/725

II. FIELDS SEARCHED

Minimum Documentation Searched²

Classification System	Classification Symbols
IPC	C08B37/00, A61K31/725, A61K35/56

Documentation Searched other than Minimum Documentation
to the Extent that such Documents are Included in the Fields Searched³

III. DOCUMENTS CONSIDERED TO BE RELEVANT⁴

Category ⁵	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
X	Chemical Abstracts No. CA109(5): 31852c (Zhongguo Yaolixue Yu Dulixue Zazhi, 2(2), 98-101 (1988) Zhang, Peiwen; Luo, Sufang; Zhong, Chunming; Wang, Qiangji: Abstract of "Anticoagulant effect of Holothuria leucospilota acid mucopolysaccharide.")	1 - 10
X	Chemical Abstracts No. CA107(21): 190638n (Zhongguo Yaoli Xuebao, 8(5), 447-50 (1987) Shan, Chunwen; Liang, Qizhao; Wang, Qiangji: Abstract of "Effects of an acidic polysaccharide from Holothuria leucospilota on the platelet aggregation in rabbits.")	1 - 10
X	Chemical Abstracts No. CA99(3): 19901k (Yaokxue Xuebao, 18(3), 203-8 (1983) Fan, Huizeng; Chen, Judi; Lu, Peihong; Hao, Xiaoge; Li, Haitang: Abstract of "Acidic polysaccharides from Holothuria leucospilota (Brandt).")	1 - 10

* Special categories of cited documents: ¹⁰

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search

April 3, 1990 (03. 04. 90)

Date of Mailing of this International Search Report

April 16, 1990 (16. 04. 90)

International Searching Authority

Japanese Patent Office

Signature of Authorized Officer

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

X	Chemical Abstracts No. CA97(3): 20682e (Yaoxue Tongbao, 16(10), 631 (1981) Fan, Huieng; Chen, Judi; Lu, Peihong; Li, Haitang: Abstract of "Acidic polysaccharides from Holothuria leucospilota.")	1 - 10
X	Chemical Abstracts No. CA94(8): 52763m (Yao Hsueh Hsueh Pao, 15(5), 263-70 (1980) Fan, Hui - Zeng; Chen, Ju - Di; Lin, Ke - Zhong: Abstract of "Isolation of an acidic mucopolysaccharide from Stichopus japonicus selenka and some of its physical and chemical properties.")	1 - 10

V. OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE ¹

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. Claim numbers because they relate to subject matter not required to be searched by this Authority, namely:

2. Claim numbers because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. Claim numbers because they are dependent claims and are not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING ²

This International Searching Authority found multiple inventions in this international application as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.
2. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:
3. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:
4. As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remark on Protest

- The additional search fees were accompanied by applicant's protest.
- No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

X	Chemical Abstracts No. CA84(25): 175525m (Chem. Pharm. Bull., 24(2), 275-84 (1976) Kitagawa, I.; Sugawara, T.; Yosioka, I.: Abstract of "Saponin and sapogenol. XV. Antifungal glycosides from the sea cucumber Stichopus japonicus Selenka.")	1 - 10
A	JP, A, 61-22021 (Nichirei Corp.), 30 January 1986 (30. 01. 86), (Family: none)	1 - 10
A	JP, A, 63-128001 (Taiho Pharmaceutical Co., Ltd. and one other), 31 May 1988 (31. 05. 88), (Family: none)	1 - 10

V. OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE¹

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